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### Review

# Memory T-cell-specific therapeutics attenuate allograft rejection via mediation of alloreactivity in memory cells

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#### 1. Introduction

Immunological memory is a fundamental hallmark of adaptive immunity and plays a critical role in providing antigen-specific protective immunity in higher vertebrates [1]. The immune repertoire is constantly being imprinted through exposure to environmental antigens, resulting in their rapid clearance following re-exposure to the same antigen [1]. In adults, 40–50% of T cells in the peripheral blood have memory phenotypes [2]. It is thought that memory T cells ( $T_Ms$ ) are generated through continuous exposure to bacterial and viral pathogens [3], blood transfusions or pregnancies, as well as transplantation [4–6]. Primed T cells take on characteristics indicative of prior activation and give rise to a population of cells collective referred to as  $T_Ms$ . These  $T_Ms$ mediate protective immune responses by invading pathogens and are thought to provide an evolutionary survival advantage [1].

#### ABSTRACT

Many means in inbred rodent models promoted long-term graft survival or donor-specific tolerance, but less so in nonhuman primates, outbred rodents or human patients. A diverse repertoire of memory T cells, derived from heterologous immunity or prior to exposure to alloantigen, has been believed to be an important part of this barrier. Memory T cells have a unique capacity to generate effector functions quickly upon re-exposure to antigen, and this capacity is achieved by reduced activation thresholds, and expressed high level trafficking and adhesion molecules, which is likely responsible for their exhibiting differential susceptibility to immune therapeutics compared with naïve T cells. This review outlines recent progress on characteristics of memory T cells and focuses on these potential therapies targeting memory T cells which are likely to ameliorate allograft rejection by inducing transplant tolerance.

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However, there is an increasing amount of evidence to indicate that  $T_Ms$  can cross-react with alloantigens, partially accounting for the high frequency of alloreactivity [7,8]. Alloreactive  $T_Ms$  can become activated by specific pathogens, or due to a molecular resemblance between a microbial antigen/self major histocompatibility complex (MHC) and cross-reactivity with an allogeneic MHC T cell [9]. Such cross-reactivity of  $T_Ms$  can lead to pre-sensitization in a potential recipient despite lack of exposure to tissues from a donor [10,11]. In the context of transplantation, emerging evidence has revealed the issues associated with  $T_Ms$  for transplant recipients. The  $T_Ms$  generate antigen-specific protective immunity, and pose a barrier to longtime allograft survival and transplant tolerance by cross-reactivity with allogeneic antigens [3]. In this review, we describe recent advances related to the characteristics of  $T_Ms$ , and discuss their roles in allograft rejection and potential therapies.

#### 2. Characteristics of T<sub>M</sub>s

Naïve T cells ( $T_Ns$ ) emerge from the thymus with a nonactivated phenotype characterized by T cell receptor (TCR) density and limited adhesion molecules. The phenotypes of  $T_Ns$  remain constant until they are primed by an antigen, at which point they are then considered  $T_Ms$  [1]. The expression of chemokine, selectin, and integrin receptors is altered after the activation of  $T_Ms$ .  $T_Ms$  also express higher levels of CD2, CD11a and CD44 as compared with their naïve counterparts. In humans,  $T_Ms$  express the RO isoform of CD45 as opposed to the RA isoform, and possess direct cytolytic functions in vivo [1]. In contrast to  $T_Ns$ ,  $T_Ms$  exhibit enhanced



Abbreviations:  $T_Ms$ , memory T cells; MHC, major histocompatibility complex;  $T_Ns$ , naïve T cells; TCR, T cell receptor;  $T_{CM}s$ , central memory T cells;  $T_{EM}s$ , effector memory T cells;  $T_Es$ , effector T cells; HLA, human leukocyte antigen; EBV, Epstein–Barr virus; CTLA4, cytotoxic T-lymphocyte antigen 4; LFA-3, leukocyte function-associated antigen-3; GVHD, graft-versus-host disease; mAb, monoclonal antibody; S1P, sphingosine 1-phosphate; JAKs, janus kinases;  $\gamma c$ ,  $\gamma$  chain; IkB, inhibitor of kappa-light-chain-enhancer of activated B; NF- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B; NC- $\kappa$ B, nuclear f

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activation, distinct phenotype trafficking, and functions related to adhesion. Upon re-exposure to an antigen, they secret cytokines faster than naïve T cells with reduced activation thresholds [12].

Emerging evidence has shown that  $T_Ns$  and  $T_Ms$  can cross-react with alloantigens, and the specificity of  $T_Ns$  and  $T_Ms$  pools is likely to be quite different [13]. The issue of whether alloreactive frequencies in  $T_Ms$  pools are higher, lower or equivalent as compared with  $T_Ns$  has been thoroughly discussed over the last two decades. Recently, research has shown that alloreactive  $T_Ms$  are likely to be limited to a few discrete TCR clones when compared with  $T_Ns$ . This is likely owing to the restricted TCR repertoires of  $T_Ms$ . However, the frequency of a given clone is higher in  $T_Ms$  pools compared with  $T_Ns$  pools [14,15]. TCR clones among  $T_Ms$  are therefore thought to be responsible for higher alloreactivity than that seen in  $T_Ns$ . In the context of transplantation, pre-existing donor-specific  $T_Ms$  are believed to participate in the development of acute and chronic rejection.

#### 2.1. T<sub>M</sub> subsets

The phenotype of heterogeneous  $T_M s$  has been classified into two subsets: central  $T_M s$  ( $T_{CM} s$ ; CD62L<sup>hi</sup>CCR7<sup>+</sup>), which circulate in non-lymphoid or lymphoid tissues and proliferate in response to stimulation when an antigen is encountered; and effector  $T_M s$ ( $T_{EM} s$ ; CD62L<sup>lo</sup>CCR7<sup>-</sup>), which migrate to non-lymphoid tissues and release cytokines thereby providing an effector function at peripheral sites [1,16,17]. This classification highlights that the overall frequency within  $T_M s$  pools may be similar, but their behavior may be distinct, depending on differentiation status.

#### 2.2. Models of $T_M$ generation

Whether  $T_Ms$  arise from naïve or effector T cells ( $T_Es$ ) remains unclear, four models have been suggested to describe the generation of  $T_Ms$ . First, a separate lineage model suggests that effector and  $T_Ms$  can be directly generated by homeostatic turnover of different naïve precursors [18–20] (Fig. 1A). Second, naïve cells undergoing their first division produced unequal numbers of effector and memory populations [21] (Fig. 1B). Third, a linear progression model postulates that memory populations are derived from a pool of primed effector cells that experienced expansion and contraction before finally becoming  $T_Ms$  [22,23] (Fig. 1C). Fourth, during the latter stages of an immune response, a remaining, small amount of antigen fails to adequately stimulate T cells; therefore, naïve T cells differentiate into memory populations [24] (Fig. 1D).

#### 3. The role of T<sub>M</sub>s in allograft rejection

Using inbred rodent models, long-term graft survival or donorspecific tolerance has been achieved several times. Success has been limited in nonhuman primates, outbred rodents, and human patients. A diverse range of T<sub>M</sub>s derived from heterologous immunity or prior exposure to alloantigens, are believed to be an important part of this barrier [3,25-27]. A study by Felix and Allen suggested that alloreactive T cells have the ability to recognize many unrelated peptide sequences [28]. Burrows et al. demonstrated that human leukocyte antigen (HLA)-B8-restricted Epstein-Barr virus (EBV)-specific T cells were cross-reactive with HLA-B14, -44, and -35 [29,30]. Therefore, alloreactive T<sub>M</sub>s could mediate allograft rejection by cross-reacting with donor antigens. It has been reported that pre-transplant frequency of donor-specific T<sub>M</sub>s correlates with the risk of post-transplant acute rejection episodes [31]. Donor-reactive T<sub>M</sub>s can migrate into allograft tissues within 24 h post-transplant [32]. Tolerance induction protocols in naïve mice based on a co-stimulatory blockade of cytotoxic Tlymphocyte antigen 4 (CTLA4-Ig) and anti-CD40L have been found to be ineffective in mice infected with lymphocytic choriomeningitis virus [33]. In another study, depletion of CD8  $T_Ms$  effectively improved induction of chimerism and renal survival [34]. Taken together, these data suggest that donor-specific  $T_Ms$  contributed to graft destruction and a resistance in the induction of tolerance.

#### 4. T<sub>M</sub>s and immunosuppressants

Allograft recipients require long-term immunosuppression to prevent acute and chronic rejection. Immunosuppressive agents act in different ways to elicit their effects. Immunosuppression can be carried out by blocking co-stimulatory signals (CTLA4-Ig); blocking TCR and cytokine receptor signaling (cyclosporine A, FK506); T cell depletion (antithymocyte globulin); mixed chimerism (bone marrow transfusion); and blocking trafficking (alefacept, FTY720). The majority of immunosuppressive agents mentioned above focus on naïve T cells. In renal transplant patients, treatment with antithymocyte globulin resulted in a 90% depletion of T<sub>M</sub>s and T<sub>N</sub>s, with CD45RO<sup>+</sup>CD62L<sup>lo</sup> T<sub>M</sub>s comprising the dominant lymphocyte population [35]. A blockade of co-stimulatory molecules (CD154/CD40 or ICOS/ICOSL) inhibited rejection response and prolonged graft survival, but not in alloantigen-primed mice [36-38]. It has been reported that using cyclosporine A alone shows a dose-dependent enhancement of memory recall [39,40]. Growing evidence has revealed that T<sub>M</sub>s exhibit different susceptibilities to immune therapeutics, and this distinguishes them from T<sub>N</sub>s. Because of this property, the potential number of immunosuppressive agents available for targeting T<sub>M</sub>s is impressive (Table 1).

#### 4.1. Effect of blocking T<sub>M</sub>s trafficking

T<sub>M</sub>s express higher levels of CD2 and CD11 compared with T<sub>N</sub>s, and can rapidly traffic into grafts and mediate rejection by initiating effector functions [41]. Thus, blockade of trafficking might be one potential immunosuppressive strategy. A candidate agent is alefacept, a dimeric, fusion protein consisting of the extracellular CD2-binding portion of human leukocyte function-associated antigen-3 (LFA-3) linked to the Fc portion of human immunoglobulin G. Several mechanisms are thought to attenuate allograft rejection including complement-mediated lysis and blocking of LFA-3 as the CD2 pathway is important in reactivation of T<sub>EM</sub>s [42,43]. Alefacept is approved for the clinical treatment of psoriasis by the FDA, and its therapeutic effects have been associated with the ability to eliminate T<sub>M</sub>s. It has been reported that alefacept prolongs traditional co-stimulatory blockades. This is based on renal allograft survival times in nonhuman primates [43]. Data acquired from Shapira et al. showed that alefacept preferentially decreases circulating T<sub>M</sub> subsets in refractory chronic extensive graft-versushost disease (GVHD) in humans [44]. In non-human primates, islet transplantation with alefacept treatment prolonged allograft survival by targeting co-stimulatory blockade-resistant T<sub>EM</sub>s [45].

Efalizumab, an anti-human leukocyte function-associated antigen-1 (LFA-1) monoclonal antibody (mAb), inhibits the interaction of LFA-1 with CD11a to interrupt  $T_M$  trafficking (Fig. 2A). It has been reported that anti-LFA-1 mAbs attenuate donor-reactive memory recall responses and reduce T-cell trafficking to the allograft in murine models of transplantation [46,47]. Phase 2 clinical trials involving renal transplantation demonstrated that anti-LFA-1 mAbs have potential inhibitory effects on  $T_Ms$  [48].

FTY720 is a high-affinity agonist for sphingosine 1-phosphate (S1P) receptors. It can inhibit lymphocyte migration from the thymus and peripheral lymphoid tissues by down-regulating surface expression of the S1P receptor [49] (Fig. 2A). FTY720 leads to peripheral blood lymphopenia within 3–24 h after administration to rats [50]. FTY720 promoted cyclosporine A-based skin, cardiac



**Fig. 1.** Four different models of  $T_M$  generation. (A)  $T_E$ s and  $T_M$ s are directly generated by homeostatic turnover of different  $T_N$ s; (B)  $T_N$ s undergoing their first division produced unequal numbers of  $T_E$ s and  $T_M$ s; (C) memory populations are derived from a pool of primed  $T_E$ s that experienced expansion and contraction; (D)  $T_N$ s differentiate into memory populations by shortage stimulation of a small amount of antigen in the latter stages of an immune response.

and renal allograft survival in rats and dogs [50–54]. Further studies have proposed that FTY720 prolongs graft survival by inhibiting alloantigen-specific CD4 memory T cells from trafficking to graft sites.

#### 4.2. Effects of blocking signaling through the cytokine receptor

In mammals, Janus kinases (JAKs) are a family of cytoplasmic tyrosine kinases consisting of four members: Jak1, Jak2, Jak3, and tyrosine kinase 2 which are predominantly expressed in hematopoietic cells and highly regulated with cell development and activation. JAKs share a common  $\gamma$  chain ( $\gamma$ c) surface receptor for IL-2, -4, -7, -9, -15, -17, and -21, which regulates the development, activation, proliferation, and survival of T, B, NK, and NKT cells [55]. Recent studies have suggested that cytokines, including IL-2, -7 and -15, play an important role in generation, maintenance and proliferation of T<sub>M</sub>s [56] (Fig. 2A). Therefore, JAKs are a potential molecular target for blockade of cytokine signals used by T<sub>M</sub>s. A highly selective and potent JAK3 inhibitor, tasocitinib (also known

## Table 1

The rapeutic agents targeting  $T_{\mbox{\scriptsize M}}s.$  as CP-690550), has been shown to prevent allograft rejection in mice subjected to heart transplants, and in non-human primates that have undergone kidney transplants [57,58].

#### 4.3. Effects of inhibiting nuclear factor

# kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ B) nuclear translocation

Bortezomib (Velcade), an FDA-approved dipeptidyl boronic acid analog, is the first proteasomal inhibitor that prevents degradation of inhibitors of  $\kappa$ B (I $\kappa$ B) and thus blocks NF- $\kappa$ B nuclear translocation [59] (Fig. 2A). It has been reported that bortezomib mediates apoptosis in alloreactive human T cells [60]. A recent study demonstrated that bortezomib can suppress the proliferation of rapamycin-resistant T<sub>M</sub>s populations in non-human primates [61]. More importantly, bortezomib was shown to preserve the function of regulatory T cells (Tregs) while inhibiting the activation of donor-reactive T<sub>M</sub>s. Thus, Bortezomib and a newer generation

Therapeutic agents	Effects on T <sub>M</sub> s	Mechanism of action	References
Alefacept	Blockade of T <sub>M</sub> s trafficking	Block leukocyte LFA-3:CD2 pathway	[35–38]
Efalizumab		Inhibit the interaction of LFA-1 and CD11a, and attenuate T <sub>M</sub> s recall responses	[39–41]
FTY720		Down-modulate of the surface expression of S1P receptor	[42-47]
Tasocitinib	Blockade of signaling through the cytokine receptor	Block the signals of cytokines such as IL-2, IL-7 and IL-15 which play an important role in generation, maintenance and proliferation of $T_Ms$	[48–51]
Bortezomib	Induction of T <sub>M</sub> s apoptosis	Prevent degradation of IкB and thus blocks NF-кB nuclear translocation	[52–54]
Tolerogenic dendritic cells	Induction of T <sub>M</sub> s tolerance	Inhibit allo-immune responses of T <sub>M</sub> s, induce anergy and hyporesponsiveness in T <sub>M</sub> s	[55,60–67]
Regulatory T cells		Inhibit allo-immune responses of T <sub>M</sub> s	[67,73]



**Fig. 2.** Therapeutic agents targeting T<sub>M</sub>s. (A) Three types of immunosuppressive agents available for targeting T<sub>M</sub>s, (I) include Alefacept, Efalizumab, and FTY720 which block T<sub>M</sub>s trafficking, (II) include Tasocitinib which blocks T<sub>M</sub>s proliferation, (III) include Bortezomib which induces T<sub>M</sub>s apoptosis; (B) Tol DCs could induce tolerance of T<sub>M</sub>s which suggested its potential to regulate allo-immune responses in transplant patients; (C) Tregs could inhibit allo-responses of T<sub>M</sub>s which may prolong graft survival.

of more selective and less toxic proteasome inhibitors are showing promise as inhibitors of donor-reactive T<sub>M</sub>s responses.

#### 5. T<sub>M</sub>s and tolerogenic dendritic cells (Tol DCs)

Overcoming the resistance of  $T_M s$  to tolerance induction continues to present a serious challenge in transplantation. Tol DCs are a promising, novel immunotherapeutic tool for inhibiting naïve T cell-mediated immune rejection [62] (Fig. 2B). The mechanisms by which Tol DCs induce tolerance include  $T_N s$  deletion, anergy, cytokine deviation, and/or the induction of Tregs [63–66]. Several recent studies have addressed the importance of Tol DCs in tolerance induction of  $T_M s$ . It was previously shown that monoclonal anti-CD3 or fixed DCs induced hyporesponsiveness in  $T_M s$  but not in  $T_N s$  [67–69]. Co-cultures of dexamethasone-conditioned human DCs induced anergy in terms of proliferation and cytokine production [70]. Tolerance induction protocols based on human Tol DCs generated with dexamethasone induced anergy in allogeneic CD4<sup>+</sup>  $T_M s$ , which retained their capacity to produce large amounts of

IL-10. Anergy in T<sub>M</sub>s did not induce Tregs and could be partially reversed by IL-2 [62]. Our data demonstrated that RelB-silenced DCs induced hyporesponsiveness in CD4 $^+$   $T_Ns$  and CD4 $^+$   $T_Ms.$  Upon re-stimulation with mature DCs, CD4 $^+$   $T_Ns$  primed with RelBsilenced DCs maintain responsiveness, while CD4<sup>+</sup> T<sub>M</sub>s primed by RelB-silenced DCs maintain hyporesponsiveness in terms of proliferation and cytokine production. This would suggest that RelB-silenced DCs induce a hyporesponsive state in CD4<sup>+</sup> T<sub>N</sub>s and CD4+  $T_Ms$  in a distinct manner [71]. CD8+  $T_Ms$  also are potent barrier to transplant tolerance induction. Research has demonstrated that CD8<sup>+</sup> T<sub>M</sub>s are susceptible to tolerance induction when cognate antigens are expressed in DCs [72]. A recently study has shown that CD4<sup>+</sup> T<sub>M</sub>s responses could be terminated by steadystate DCs expressing cognate antigen [73]. Plasmacytoid dendritic cells (pDCs) are associated with tolerance to allografts in experimental animal models, and adoptive transfusion of donor-derived pDCs significantly prolonged allogeneic heart graft and skin graft survival. Moreover, pDCs could suppress responses in unfractionated allogeneic  $CD_3^+$  T cells that contain both  $T_N$ s. and  $T_M$ s [74]. The important ability of Tol DCs to induce tolerance of  $T_M s$  underscores the potential of these cells to regulate allo-immune responses in transplant patients.

#### 6. T<sub>M</sub>s and Tregs

Tregs are a subpopulation of T cells, defined by their role in inhibiting immune reactions and inducing tolerance [75]. Tregs play a crucial role in maintaining donor-specific hyporesponsiveness in renal and liver transplant patients [76-78]. In rodent models, adoptive transfer of Tregs prevents allograft rejection and prolongs survival of grafts [79] (Fig. 2C), suggesting that Tregs could function as a potential therapy by inhibiting T<sub>N</sub>s. Because of the resistance of T<sub>M</sub>s immunomodulation, researchers have addressed the impact of Tregs on T<sub>M</sub>s. Anti-OX40L prolonged graft survival by inhibition and apoptosis of T<sub>M</sub>s in pre-sensitized recipients. Anti-OX40L, LF-15-0195, and anti-CD45RB restored heart allograft tolerance in pre-sensitized model by induction of Tregs. However, following CD25<sup>+</sup> T-cell depletion, heart grafts were rapidly rejected, despite pre-sensitized recipients being co-administered three immunosuppressants [80]. This indicates that Tregs targeting T<sub>M</sub>s are crucial for tolerance induction in pre-sensitized heart transplant recipients. CD8<sup>+</sup> Tregs induced by pDCs suppressed the allo-responsiveness of T<sub>M</sub>s [74]. These finding are highly relevant to clinical transplantation, and support the use of Tregs as a potential approach to induce tolerance of recipient allo-reactive T<sub>M</sub>s.

#### 7. Conclusion

Over the past decade, studies published have definitively demonstrated that T<sub>M</sub>s pose a potent barrier in obtaining long-term allograft survival and transplant tolerance because heterogeneous T<sub>M</sub>s develop different susceptibilities to traditional immunosuppressive agents. In this study, we summarized two strategies for immunosuppressive and cellular therapy. Immunosuppressants can be divided into three types according to their different effects on T<sub>M</sub>s. Cellular therapies mainly use two cell types: Tol DCs and Tregs. Some immunosuppressants are already used clinically. However, because their effects are mainly based on inhibition of T<sub>M</sub> activation and proliferation, there can be many side effects on transplant patients, thereby limiting their extensive use. Because the aim of cellular therapy is to induce tolerance of T<sub>M</sub>s, leading to anergy and hyporesponsiveness of T<sub>M</sub>s to transplant allografts, it is more likely to have fewer side effects and maintain allograft functions. Although most cellular therapies are still in the early stages, some animal studies have already showed great potential in preventing rejection, prolonging survival time, and maintaining the functions of allografts. We believe that cellular therapy will become the preferred solution for clinical transplant rejection in the future.

In summary, with advances in understanding the mechanisms of  $T_M$  generation, activation, function, or trafficking, and the identification of biomarkers, potential approaches targeting  $T_M s$  should eliminate or attenuate allograft rejection.

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#### References

- Andrew JP, Mandy LF, Allan DK. Memory T-cell-specific therapeutics in organ transplantation. Curr Opin Organ Transplant 2009;12:643–9.
- [2] McFarland RD, Douek DC, Koup RA, Picker LJ. Identification of a human recent thymic emigraft phenotype. Proc Natl Acad Sci USA 2000;97:4215–20.

- [3] Ford ML, Kirk AD, Larsen CP. Donor-reactive T-cell stimulation history and precursor frequency: barriers to tolerance induction. Transplantation 2009;87: S69–74.
- [4] Kaminski ER, Hows JM, Goldman JM, Batchelor JR. Lymphocytes from multitransfused patients exhibit cytotoxicity against autologous cells. Br J Haematol 1992;81(1):23–6.
- [5] van Kampen CA, Versteeg-van der Voort Maarschalk MF, Langerak-Langerak J, van Beelen E, Roelen DL, Claas FH. Pregnancy can induce long-persisting primed CTLs specific for inherited paternal HLA antigens. Hum Immunol 2001;62(3):201–7.
- [6] Pelletier RP, Hennessy PK, Adams PW, Orosz CG. High incidence of donorreactive delayed-type hypersensitivity reactivity in transplant patients. Am J Transplant 2002;2(10):926–33.
- [7] Lombardi G, Sidhu S, Daly M, Batchelor JR, Makgoba W, Lechler RI. Are prime alloresponses truly primary? Int Immunol 1990;2(1):9–13.
- [8] Lechler RI, Heaton T, Barber L, Bal V, Batchelor JR, Lombardi G. Molecular mimicry by major histocompatibility complex molecules and peptides account for some alloresponses. Immunol Lett 1992;34(1):63–9.
- [9] Valujskikh A, Baldwin 3rd WM, Fairchild RL Recent progress and new perspectives in studying T cell responses to allografts. Am J Transplant 2010;10(5):1117–25.
- [10] Deacock SJ, Lechler RI. Positive correlation of T cell sensitization with frequencies of alloreactive T helper cells in chronic renal failure patients. Transplantation 1992;54(2):338–43.
- [11] Padovan E, Casorati G, Dellabona P, Giachino C, Lanzavecchia A. Dual receptor T-cell. Implications for alloreactivity and autoimmunity. Ann N Y Acad Sci 1995;756:66–70.
- [12] Farber DL. Biochemical signaling pathways for memory T cell recall. Semin Immunol 2009;9:89–97.
- [13] Ford ML, Larsen CP. Overcoming the memory barrier in tolerance induction: molecular mimicry and functional heterogeneity among pathogen-specific Tcell populations. Curr Opin Organ Transplant 2010;15(4):405–10.
- [14] Sourdive DJ, Murali-Krishna K, Altman JD, Zajac AJ, Whitmire JK, Pannetier C, et al. Conserved T cell receptor repertoire in primary and memory CD8 T cell responses to an acute viral infection. J Exp Med 1998;188(1):71–82.
- [15] van Leeuwen EM, Remmerswaal EB, Heemskerk MH, ten Berge IJ, van Lier RA. Strong selection of virus-specific cytotoxic CD4<sup>+</sup> T-cell clones during primary human cytomegalovirus infection. Blood 2006;108(9):3121–7.
- [16] Sallusto F, Lenig D, Forster R, Lipp M, Lanzavecchia A. Two subsets of memory T lymphocytes with distinct homing potentials and effector functions. Nature 1999;401(6754):708–12.
- [17] Seder RA, Ahmed R. Similarities and differences in CD4<sup>+</sup> and CD8<sup>+</sup> effector and memory T cell generation. Nat Immunol 2003;4(9):835–42.
- [18] Goldrath AW, Bogatzki LY, Bevan MJ. Naive T cells transiently acquire a memory-like phenotype during homeostasis-driven proliferation. J Exp Med 2000;192:557–64.
- [19] Kieper WC, Jameson SC. Homeostatic expansion and phenotypic conversion of naive T cells in response to self peptide/MHC ligands. Proc Natl Acad Sci USA 1999;96:13306–11.
- [20] Ge Q, Hu H, Eisen HN, Chen JZ. Different contributions of thymopoiesis and homeostasis-driven proliferation to the reconstitution of naive and memory T cell compartments. Proc Natl Acad Sci USA 2002;99:2989–94.
- [21] Chang JT, Palanivel VR, Kinjyo I, Schambach F, Intlekofer AM, Banerjee A, et al. Asymmetric T lymphocyte division in the initiation of adaptive immune responses. Science 2007;315(5819):1687–91.
- [22] Lohning M, Hegazy AN, Pinschewer DD, Busse D, Lang KS, Hofer T, et al. Longlived virus-reactive memory T cells generated from purified cytokine-secreting T helper type 1 and type 2 effectors. J Exp Med 2008;205(1):53–61.
- [23] Harrington LE, Janowski KM, Oliver JR, Zajac AJ, Weaver CT. Memory CD4 T cells emerge from effector T-cell progenitors. Nature 2008;452(7185): 356–60.
- [24] Moulton VR, Bushar ND, Leeser DB, Patke DS, Farber DL. Divergent generation of heterogeneous memory CD4 T cells. J Immunol 2006;177:869–76.
- [25] Preston EH, Xu H, Dhanireddy KK, Pearl JP, Leopardi FV, Starost MF, et al. IDEC-131 (anti-CD154), sirolimus and donor-specific transfusion facilitate operational tolerance in non-human primates. Am J Transplant 2005;5:1032–41.
- [26] Adams AB, Pearson TC, Larsen CP. Heterologous immunity: an overlooked barrier to tolerance. Immunol Rev 2003;196:147–60.
- [27] Brook MO, Wood KJ, Jones ND. The impact of memory T cells on rejection and the induction of tolerance. Transplantation 2006;82:1–9.
- [28] Felix NJ, Allen PM. Specificity of T-cell alloreactivity. Nat Rev Immunol 2007;7:942–53.
- [29] Burrows SR, Silins SL, Moss DJ, Khanna R, Misko IS, Argaet VP. T cell receptor repertoire for a viral epitope in humans is diversified by tolerance to a background major histocompatibility complex antigen. J Exp Med 1995;182:1703–15.
- [30] Burrows SR, Silins SL, Khanna R, Burrows JM, Rischmueller M, McCluskey J, et al. Cross-reactive memory T cells for Epstein–Barr virus augment the alloresponse to common human leukocyte antigens: degenerate recognition of major histocompatibility complex-bound peptide by T cells and its role in alloreactivity. Eur J Immunol 1997;27:1726–36.
- [31] Peter SH, Neil SG, Shannon K, Cora D, Donald EH, James AS, et al. Pretransplant frequency of donor-specific IFN-{gamma}-producing lymphocytes is a manifestation of immunologic memory and correlates with the risk of posttransplant rejection episodes. J Immunol 1999;163:2267–75.

- [32] Schenk AD, Nozaki T, Rabant M, Valujskikh A, Fairchild RL. Donor-reactive CD8 memory T cells infiltrate cardiac allografts within 24 hours post-transplant in naive recipients. Am | Transplant 2008;8(8):1652–61.
- [33] Adams AB, Williams MA, Jones TR, Shirasuqi N, Durham MM, Kaech SM, et al. Heterologous immunity provides a potent barrier to transplantation tolerance. | Clin Invest 2003;111:1887–95.
- [34] Koyama I, Nadazdin O, Boskovic S, Ochiai T, Smith RN, Sykes M, et al. Depletion of CD8 memory T cells for induction of tolerance of a previously transplanted kidney allograft. Am J Transplant 2007;7(5):1055–61.
- [35] Pearl JP, Parris J, Hale DA, Hoffmann SC, Bernstein WB, McCoy KL, et al. Immunocompetent T-cells with a memory-like phenotype are the dominant cell type following antibody-mediate T-cell depletion. Am J Transplant 2005;5:465–74.
- [36] Zhai Y, Meng L, Gao F, Busuttil RW, Kupiec-Weqlinski JW. Allograft rejection by primed/memory CD8<sup>+</sup> T cells is CD154 blockade resistant: therapeutic implications for sensitized transplant recipients. J Immunol 2002;169:4667–73.
- [37] Chen Y, Heeger PS, Valujskikh A. In vivo helper functions of alloreactive memory CD4b T cells remain intact despite donor-specific transfusion and anti-CD40 ligand therapy. J Immunol 2004;172:5456–66.
- [38] Kashizuka H, Sho M, Nomi T, Ikeda N, Kuzumoto Y, Akashi S, et al. Role of the ICOS-B7h costimulatory pathway in the pathophysiology of chronic allograft rejection. Transplantation 2005;79:1045–50.
- [39] Bemer V, Motta I, Perret R, Truffa-Bachi P. Interleukin-2 down-modulates memory Thelper lymphocyte development during antigenic stimulation in vitro. Eur J Immunol 1995;25:3394–401.
- [40] Gupta M, Satyaraj E, Durdik JM, Rath S, Bal V. Differential regulation of T cell activation for primary versus secondary proliferative responses. J Immunol 1997;158:4113–21.
- [41] Andrew JP, Mandy LF, Allan DK. Memory T-cell-specific therapeutics in organ transplantation. Curr Opin Organ Transplant 2009;14:643–9.
- [42] Michael PH, Craig LL. Alefacept for psoriasis. Semin Cutan Med Surg 2010;29(1):53-5.
- [43] Weaver TA, Charafeddine AH, Agarwal A, Turner AP, Russell M, Leopardi FV, et al. Alefacept promotes costimulation blockade based allograft survival in nonhuman primates. Nat Med 2009;15:746–9.
- [44] Shapira MY, Abdul-Hai A, Resnick IB, Bitan M, Tsirigotis P, Gesundheit B, et al. Alefacept treatment for refractory chronic extensive GVHD. Bone Marrow Transplant 2009;43:339–43.
- [45] Turner AP, Weaver T, Badell R, Cano J, Avila J, Turner P, et al. LFA-3-Ig (Alefacept) prolongs allograft survival by targeting costimulation blockade resistant T effector memory cells in non-human primate islet transplantation. Am J Transplant 2009;9(Suppl. 2):237.
- [46] Ford M, Wegener M, Cooper S, Kirk A, Larsen C. Overcoming the memory barrier: role of anti-LFA-1 in promoting long-term graft survival in recipients possessing donor-reactive memory T cells. Am J Transplant 2009;9(Suppl. 2):536.
- [47] Sétoguchi K, Schenk A, Ishii D, Tanabe K, Fairchild R. Anti-lymphocyte function associated antigen-1 (LFA-1) monoclonal antibody prevents graft-infiltrating CD8<sup>+</sup> memory cells early post-transplant. Am J Transplant 2009;9(Suppl. 2):209.
- [48] Vincenti F, Mendez R, Pescovitz M, Rajagopalan PR, Wilkinson AH, Butt K, et al. A Phase I/II randomized open-label multicenter trial of efalizumab, a humanized anti CD11a, anti-LFA-1 in renal transplantation. Am J Transplant 2007;7:1770–7.
- [49] Chiba K, Yanagawa Y, Masubuchi Y, Kataoka H, Kawaguchi T, Ohtsuki M, et al. FTY720, a novel immunosuppressant, induces sequestration of circulating mature lymphocytes by acceleration of lymphocyte homing in rats. I. FTY720 selectively decreases the number of circulating mature lymphocytes by acceleration of lymphocyte homing. J Immunol 1998;160:5037–44.
- [50] Chiba K, Hoshino Y, Suzuki C, Masubuchi Y, Yanagawa Y, Ohtsuki M, et al. FTY720, a novel immunosuppressant possessing unique mechanisms. I. Prolongation of skin allograft survival and synergistic effect in combination with cyclosporine in rats. Transplant Proc 1996;28(2):1056–9.
- [51] Hoshino Y, Suzuki C, Ohtsuki M, Masubuchi Y, Amano Y, Chiba K. FTY720, a novel immunosuppressant possessing unique mechanisms. II. Long-term graft survival induction in rat heterotopic cardiac allografts and synergistic effect in combination with cyclosporin A. Transplant Proc 1996;28(2):1060–1.
- [52] Chiba K, Yanagawa Y, Masubuchi Y, Kataoka H, Kawaguchi T, Ohtsuki M, et al. FTY720, a novel immunosuppressant, induces the sequestration of circulating mature lymphocytes by acceleration of lymphocyte homing in rats. I. FTY720 selectively decreases the number of circulating mature lymphocytes by acceleration of lymphocyte homing. J Immunol 1998;160(10):5037–44.
- [53] Kawaguchi T, Hoshino Y, Rahman F, Amano Y, Higashi H, Kataoka H, et al. FTY720, a novel immunosuppressant possessing unique mechanisms. III. Synergistic prolongation of canine renal allograft survival in combination with cyclosporin A. Transplant Proc 1996;28:1062–3.
- [54] Suzuki S, Enosawa S, Kakefuda T, Shinomiya T, Amari M, Naoe S, et al. A novel immunosuppressant, FTY720, with a unique mechanism of action, induces long-term graft acceptance in rat and dog. Transplantation 1996;61(2): 200–5.

- [55] Rochman Y, Spolski R, Leonard WJ. New insights into the regulation of T cells by gamma(c) family cytokines. Nat Rev Immunol 2009;9(7):480–90.
- [56] Schluns KS, Williams K, Ma A, Zheng XX, Lefrancois L. Cutting edge: requirement for IL-15 in the generation of primary and memory antigen-specific CD8 T cells. J Immunol 2002;168:4827–31.
- [57] Borie DC, Changelian PS, Larson MJ, Si MS, Paniagua R, Higgins JP, et al. Immunosuppression by the JAK3 inhibitor CP-690, 550 delays rejection and significantly prolongs kidney allograft survival in nonhuman primates. Transplantation 2005;79:791–801.
- [58] Changelian PS, Flanagan ME, Ball DJ, Kent CR, Magnuson KS, Martin WH, et al. Prevention of organ allograft rejection by a specific Janus kinase 3 inhibitor. Science 2003;302:875–8.
- [59] Elliott PJ, Zollner TM, Boehncke WH. Proteasome inhibition: a new antiinflammatory strategy. J Mol Med 2003;81(4):235–45.
- [60] Blanco B, Perez-Simon JA, Sanchez-Abarca LI, Carvajal-Vergara X, Mateos J, Vidriales B, et al. Bortezomib induces selective depletion of alloreactive T lymphocytes and decreases the production of Th1 cytokines. Blood 2006;107(9):3575–83.
- [61] Kim JS, Lee JI, Shin JY, Kim SY, Shin JS, Lim JH, et al. Bortezomib can suppress activation of rapamycin-resistant memory T cells without affecting regulatory T-cell viability in nonhuman primates. Transplantation 2009;88:1349–59.
- [62] Anderson AE, Sayers BL, Haniffa MA, Swan DJ, Diboll J, Wang XN, et al. Differential regulation of naive and memory CD4<sup>+</sup>T cells by alternatively activated dendritic cells. J Leukoc Biol 2008;84:124–33.
- [63] Dong X, Bachman LA, Kumar R, Griffin MD. Generation of antigen-specific, interleukin-10-producing T-cells using dendritic cell stimulation and steroid hormone conditioning. Transpl Immunol 2003;11:323–33.
- [64] Wang Z, Larregina AT, Shufesky WJ, Perone MJ, Montecalvo A, Zahorchak AF, et al. Use of the inhibitory effect of apoptotic cells on dendritic cells for graft survival via T-cell deletion and regulatory T cells. Am J Transplant 2006;6:1297–311.
- [65] Buckland M, Jago CB, Fazekasova H, Scott K, Tan PH, George AJ, et al. Aspirintreated human DCs up-regulate ILT-3 and induce hyporesponsiveness and regulatory activity in responder T cells. Am J Transplant 2006;6:2046–59.
- [66] Yates SF, Paterson AM, Nolan KF, Cobbold SP, Saunders NJ, Waldmann H, et al. Induction of regulatory T cells and dominant tolerance by dendritic cells incapable of full activation. J Immunol 2007;179:967–76.
- [67] Davis LS, Lipsky PE. Tolerance induction of human CD4<sup>+</sup>T cells: markedly enhanced sensitivity of memory versus naive T cells to peripheral anergy. Cell Immunol 1993;146:351–61.
- [68] Hayashi RJ, Loh DY, Kanagawa O, Wang F. Differences between responses of naive and activated T cells to anergy induction. J Immunol 1998;160:33–8.
- [69] Andris F, Denanglaire S, de Mattia F, Urbain J, Leo O. Naïve T cells are resistant to anergy induction by anti-CD3 antibodies. J Immunol 2004;173:3201–8.
- [70] Woltman AM, van der Kooij SW, de Fijter JW, van Kooten C. Maturationresistant dendritic cells induce hyporesponsiveness in alloreactive CD45RA<sup>+</sup> and CD45R0<sup>+</sup> T-cell populations. Am J Transplant 2006;6:2580–91.
- [71] Luo L, Li C, Wu W, Lu J, Shan J, Li S, et al. Dendritic cells transduced with lentiviral vector targeting RelB gene using RNA interference induce hyporesponsiveness in memory CD4<sup>+</sup> T cells and naive CD4<sup>+</sup> T cells. Cell Immunol 2012;273: 85–93.
- [72] Kenna TJ, Thomas R, Steptoe RJ. Steady-state dendritic cells expressing cognate antigen terminate memory CD8<sup>+</sup>T-cell responses. Blood 2008;111: 2091–100.
- [73] Nasreen M, Waldie TM, Dixon CM, Steptoe RJ. Steady-state antigen-expressing dendritic cells terminate CD4<sup>+</sup> memory T-cell responses. Eur J Immunol 2010;40:2016–25.
- [74] Boor PP, Metselaar HJ, Jonge S, Mancham S, van der Laan LJ, Kwekkeboom J. Human plasmacytoid dendritic cells induce CD8<sup>+</sup> LAG-31Foxp31CTLA-41 regulatory T cells that suppress allo-reactive memory T cells. Eur J Immunol 2011;41:1663–74.
- [75] Shan J, Guo Y, Luo L, Lu J, Li C, Zhang C, et al. Do CD4<sup>+</sup> Foxp3<sup>+</sup> Treg cells correlate with transplant outcomes: a systematic review on recipients of solid organ transplantation. Cell Immunol 2011;270:5–12.
- [76] Baan CC, Velthuis JH, van Gurp EA, Mol WM, Klepper M, Ijzermans JN, et al. Functional CD25bright+ alloresponsive T cells in fully immunosuppressed renal allograft recipients. Clinical 2007;21:63–71.
- [77] Hendrikx TK, van Gurp EA, Sewgobind VD, Mol WM, Schoordijk W, Klepper M, et al. Generation of donor-specific regulatory T-cell function in kidney transplant patients. Transplantation 2009;87(3):376–83.
- [78] Yoshizawa A, Ito A, Li Y, Koshiba T, Sakaguchi S, Wood KJ, et al. The roles of CD25<sup>+</sup>CD4<sup>+</sup> regulatory T cells in operational tolerance after living donor liver transplantation. Transplant Proc 2005;37:37–9.
- [79] Lee 4th MK, Moore DJ, Jarrett BP, Lian MM, Deng S, Huang X, et al. Promotion of allograft survival by CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells: evidence for in vivo inhibition of effector cell proliferation. J Immunol 2004;172:6539–44.
- [80] Ge W, Jiang J, Liu W, Lian D, Saito A, Garcia B, et al. Regulatory T cells are critical to tolerance induction in presensitized mouse transplant recipients through targeting memory T cells. Am J Transplant 2010;10(8):1760–73.